BLOOD PARASITES OF INDIAN TOADS

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(With 2 Figures and 2 Plates)

Introduction

Though the amphibians are known to host a large number of parasites belonging to different phyla, little attention was given to study the haematozoan parasites of amphibians. During the early nineteenth century some stray reports are available viz., Shortt (1917) and Wenyon (1926) who worked on the trypanosomes and haemogregarines of Indian toads. Recently Ray (1979a, b, 1980, 1982), Tiwari & Ray (1981) and Sinha (1979) published some papers on the blood parasites of Indian toads. No consolidated work has even been undertaken on this particular aspect. In this context, a brief account on the blood parasites known so far from the Indian toads is being dealt with in the present communication.

MATERIAL AND METHODS

Four hundred and seventy eight examples of toads comprising of 4 species viz., Bufo melanostictus, B. andersoni, B. stomaticus and B. himalayanus were collected from different parts of India and examined for blood parasites. Peripheral blood was obtained from the finger tips on alternate days and at autopsy. 'Impression' and 'Spread' preparations were made from liver, lungs, kidney and bone-marrow. Air-dried blood films and organ imprints were fixed in 100% methanol and stained with Romanowsky type of stains. For microtome section the tissue of lung, liver, kidney and spleen were fixed in Bouin's fixative, followed by general histological techniques by Pearse (1960), and stained with iron-hematoxylin and eosin.

Measurements were obtained from the camera-lucida drawing drawn on a graph paper (mm division) by counting the squares covered. The

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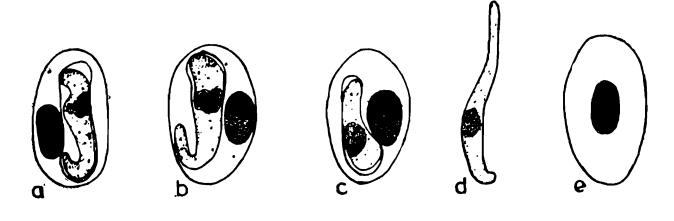
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photomicrographs were taken with the help of "Ergavel C. Z." Microscope using Olympus PM 6 attachment camera.

OBSERVATIONS

During the study on anuran haematozoa the author observed 5 intracellular parasites viz., Haemogregarina nucleobisecans, Lankesterella bufonis, Pirhemocyton sp., Cytamoeba bacterifera, Toddia sp. and 3 extracellular parasites viz., Trypanosoma rotatorium, T. chattoni and Microfilaria which have been enumerated in Table—I.

Among the parasites dealt with so far *Pirhemocyton* sp. and a sheathed microfilaria are recorded for the first time from Indian toads. Moreover,



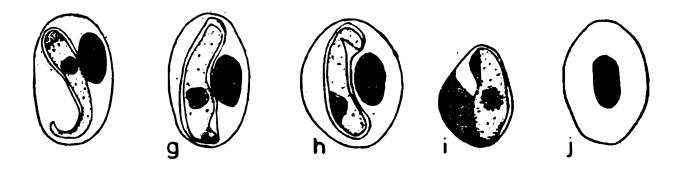


FIG. I. HAEMOGREGARINA NUCLEOBISECANS.

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Fig. 1. Camera-lucida drawings of the erythrocytic stages of Haemogregarina nucleobisecans from two anuran hosts.

a-d, the gametocyte stages in Bufo melanostictus: a & b, the longate mature form; c, the young form; d, the free gametocyte; e; the uninfected erythrocyte.

f-i, the gametocyte stages in Bufo andersoni; j, the uninfected erythrocyte.

some interesting observations on those parasites have also been made. In view of this those are discussed below:

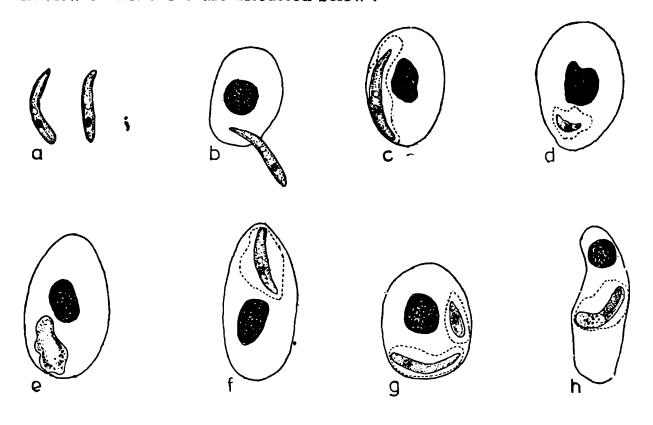


FIG.2 LANKESTERELLA BUFONIS.

TOAIM

Fig. 2. Camera-lucida drawings of Lankesterella bufonis from Bufo melanostictus.

a, the extracorpuscular sporozoites; b, the sporozoite invading the erythrocyte; c-h, the intracorpuscular sporozoites; g, the hypertrophied cell with double infection; h, the infected R. B. C. became elongate with apical displacement of the cell nucleus.

Pirhemocyton sp.

(Plate IV, Figs. 10, 11)

Hosts: Bufo melanostictus & B. himalayanus.

Description: These are intracorpuscular small rounded bodies measuring 1.5-2.0 μ m in diameter. There is a round, central chromatin dot surrounded by thin layer of cytoplasm which sometimes becomes non-visible because the enlarged chromatin meterial covers the body. There may be 2-3 small associated unstained refractile globules but not always present. These globules when present, are 1-2 μ m in diameter. There is no remarkable change in the infected erythrocytes.

Locality: Balitha, Bankura district and Darjeeling, West Bengal.

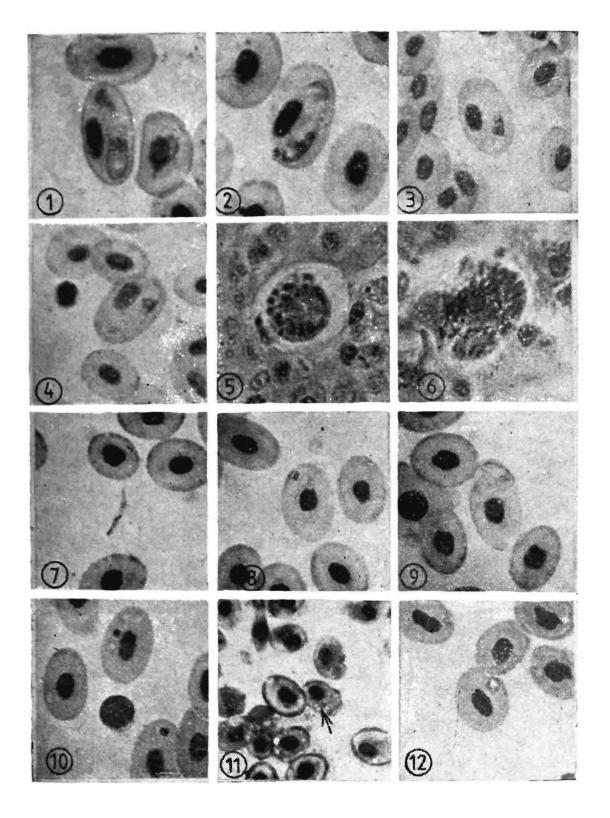
Occurrence: 12, out of 343 examples of B. melanostictus and 4 out of 32 B. himalayanus were positive for Pirhemocyton.

TABLE-1

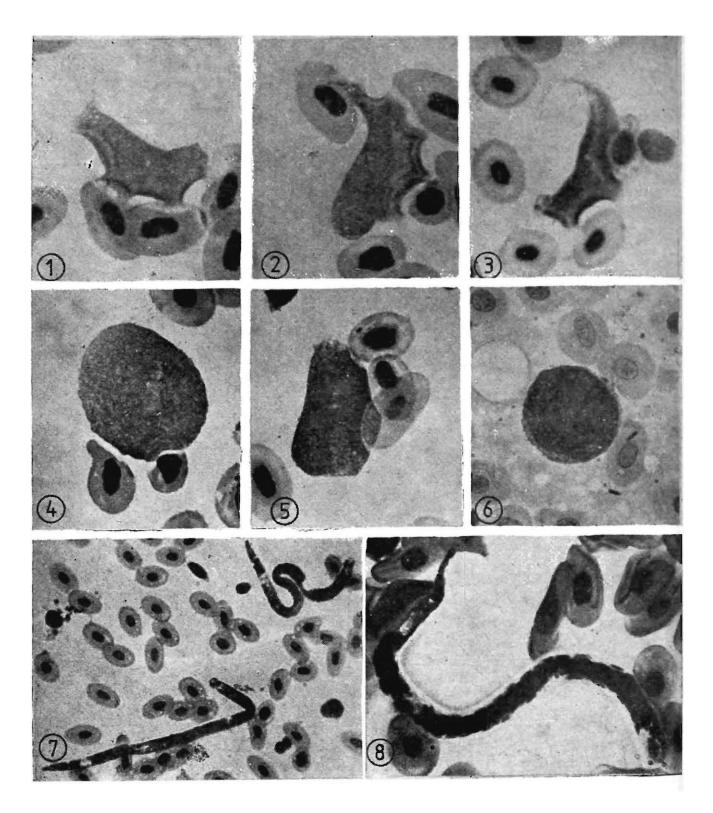
	Parasite	Name of the hosts	Locality	Diagnostic characters	Remarks	References
1.	Haemogregarina nucleobisecans (Text-figs. la-j; Pl.1, figs. 1-6)	Bufo melanostictus and B. andersoni	Bankura & Purulia District West Bengal, India.	Sausage-shaped endoglobular parasites of erythrocytes; gametocytes measuring $14.4 \times 2.95 \mu m$ with an average area of $28.95 \mu m^2$; schizogony in the hepatic cells of the liver which produces merozoites.	This species has also been recorded from a new host, Bufo andersoni.	Short t (1917), Wen yon (1926), Ray (1979b,1982).
2.	Lankesterella bufonis (Text-figs. 2a-g; Pl. 1, figs. 7-9)	Bufo melanostictus	Balitha and Kotalpur, Bankura District, West Bengal, India.	Very small, slender, crescent-shaped parasites measuring 9-11.5 μ m in length and 1.0-2.5 μ m in breadth with an average of 10.6 μ m × 1.45 μ m; nucleus terminal or subterminal and consists of few chromatin granules; a single vacuole situated centrally.	The report of this parasite represents a new host-parasite record from a new geographical region (Oriental realm).	Moham med (1962), Ray (1979b)
3.	Pirhemocyton sp. (Pl. 1, figs. 10, 11)	Bufo melanostictus & B. himalayanus	Balitha, Bankura District and Darjeeling, West Bengal, India.	Intracellular, small rounded bodies measuring 1.5-2.0 μ m in diameter; central chromatin dot surrounded by thin layer of cytoplasm; 2-3 small associated refractile globules may be present.	This parasite has been recorded from two new hosts viz. B. melanostictus and B. himalayanus.	Ray (1979b)
4.	Cytamoeba bacterifera (Pl. 1, fig. 12)	Bufo melanostictus & B. himalayanus	Balitha, Bankura Districts and Darjeeling, West Bengal, India.	Small (3-5 μ m in diam.), amoeboid or spherical, intracorpuscular red staining bodies; cytoplasm homogeneous or granular and contains crystalline bacillus-like bodies.	This parasite has been recorded for the first time from Indian toads.	Labbé (1894), Ray (19 79b).
5.	Toddia sp.	Bufo melanostictus and Bufo andersoni	Burdwan & Purulia, West Bengal, India.	Red staining spherical structure (1-3 µm in diameter); intracellular; rods of a crystaline nature associated with the parasite.	This parasite has been recorded for the first time from the toad of West Bengal.	

TABLE—1 (Concluded)

	Parasite	Name of the hosts	Locality	Diagnostic characters	Remarks	References
6.	Trypanosoma (Pl. II, figs. 1-3)	Bufo melanostictus, B. stomaticus, B. andersoni.	Balitha, Bankura District ; Purulia, West Bengal ; Bondla, Mollem & Volpoi, Goa, India.	Pleomorphic haemoflagellate occurring in 4 Distinct forms; body leaf-like, flattened and thinned out along the convex edge to form undulating membrane; cytoplasm densely granular with striated myonemes; kinetoplast dot-like and situated nearer to the posterior end; nucleus oval or round and situated on the posterior part of the body; free flagellum long (11-16 \(mm\)).		Mayer (1843) Ray (1979a, b, 1980), Ray & Choudhury (1983).
7.	Trypanosoma chattoni (Pl. II, figs. 4-6)	Bufo melanostictus, B. stomaticus, B. andersoni.	Volpoi, Mollem, Bondla, Goa, Jalpaiguri, West Bengal, India.	Rounded, spherical or irregular body: cytoplasm contains innumerable metachromatic granules and fine vacuoles; nucleus circular and central; kinetoplast very small, circular or elliptical and superimposed upon the surface of the nucleus; flagellum very short either straight or curved and extended to a certain length.	T. chattoni has been recorded from a number of bufonid hosts.	Mathis & Léger (1911), Ray (1979b, 1980), Ray & Choudhury (1983).
8.	Microfilaria (larval nematode) (Pl. II, fig. 8)	Bufo melanostictus	Amgram, 24-Parganas, West Bengal, India.	,,,,	This sheathed microfilaria is recorded for the first time in toads from India.	Ray (1979b)
9.	Microfilaria (larval nematode) (Pl. II, fig. 7)	Bufo melanostictus	Jhargram, Midnapore, West Bengal, India.	Snake-like; unsheathed, measuring 98-125 μ m in length and 5-6 μ m in width.	_	Nandi and Mandal (1977) Ray (1979b)



Figs. 1 & 2. Mature gametocytes of Haemogregarina nucleobisecans in the peri-X 1665 pheral blood of Bufo melanostictus Mature gametocytes of the same haemogregarine in the blood of Figs. 3 & 4. X 1410 Bufo andersoni A multinucleated mature schizont in the liver parenchyma of B. melanostictus X 1100 Fig. 5. developing merozoites in the liver Fig. 6. A mature schizont with X 1660 parenelyma Fig. 7. Free sporozoite of Lankesterella bufonis the blood of B. melanostictus X 1130 X 1180 Figs. 8 & 9. Intracorpuscular sporozoites of L. bufonis X 1000 Fig. 10. Pirhemocyton in Bufo melanosticius shows Pirhemocyton in the blood of Bufo Fig. 11. The arrow the X 1000 hemalayanus X 1000 Fig. 12. Cytamoeba bacterifera in the blood of Bufo melanostictus



Trypanosoma rotatorium in the peripheral blood of Bufo melanosticius Fig. 1. X 1465 Fig. 2. The same haemoflagellate in B. stomaticus X 1225 Fig. 3. T. rotatorium in B. andersoni X 1200 Fig. 4. Trypanosoma chattoni in the peripheral blood of B. melanostictus X 1225 Fig. 5. T. chattoni in B. stomaticus X 1220 Fig. 6. T. chattoni in B. andersoni X 1200 Photomicrographs of unsheathed microfilaria in the blood of Fig. 7. X 545 B. melanostictus X 835 Fig. 8. A sheathed microfilaria in the peripheral blood of B. melanostictus

Remarks: The taxonomic status of Pirhemocyton is doubtful even today. Previously it was grouped with the protozoa. Levine (1978, Personal Communication) opined that Pirhemocyton are virus inclusion bodies. Stehbens and Johnston (1966) conducted an electron microscopic studies to know the morphology of the parasite. Soltz (1971) included Pirhemocyton among a group termed as icosahedral cytoplasmic deoxyriboviruses (ICDV.). In absence of the proper knowledge about the biology of these parasites no name has been proposed.

Microfilaria (the larval nematode)

Host: Bufo melanostictus.

Sheathed form

(Plate V, Fig. 8)

Description: N=20. These microfilariae are sheathed, long slender measuring 95-120 μ m in length and 5.5-6.5 μ m in width. The sheath is hyaline, measuring 1.5-20 μ m in width from the body and often found free at the caudal end. The cephalic space is very small measuring 2.0 μ m in length. At the anterior end there are two rows of nuclei, 3 in each row and a gap in between them is well marked. The nuclear column consists of 4-5 rows of closely packed nuclei, more compact at the anterior half. The nuclei stained pink with Leishman's stain. The last nucleus is more or less rounded and located at the tip of the tail. The tail is 10.5-12.2 μ m in length, gradually narrowed and ending in a rounded tip. Position of the land marks: nerve ring 25.4-26.0%; excretory pore 34.8-35.5%; excretory cell 44.6-45.5%; inner body 69.1-70.5%; anus 82.8-84.5%.

Habitat: Blood.

Locality: Amgram, 24, Parganas district, West Bengal.

Occurrence: In one toad out of 343 examined.

Remarke: Nandi and Mandal (1977) reported one unsheathed microfilaria from Bufo melanostictus for the first time in India. In the present investigation the author also observed an unsheathed microfilaria (Fig. 7) which well corroborates with Nandi & Mandal (1977) in all respect. Therefore, the sheathed microfilaria in B. melanostictus is recorded for the first time from India. The specific determination of this larval nematode is left without finding adults.

SUMMARY

During the survey on amphibian haematozoa, 478 examples of toads comprising 4 species viz., Bufo melanostictus, B. andersoni, B. stomaticus

and B. himalayanus were examined of which the first 3 anuran hosts show the infection of Trypanosoma rotatorium (Mayer, 1843) and T. chattoni Mathis and Léger, 1911 in their blood. The blood and tissue stages of Haemogregarina nucleobisecans Shortt, 1917 have also been observed in B. melanostictus and B. andersoni. A haemococcidia viz., Lankesterella bufonis Mansour and Mohammed, 1962 has been recorded in a new host B. melanostictus for the first time from Indian subregion. Two microfilariae both sheathed and unsheathed have also been observed in the blood of B. melanostictus. Besides, some organisms of uncertain taxonomy viz., Pirhemocyton, Toddia and Cytamoeba have been found to infect B. melanostictus, B. andersoni and B. himalayanus. All those parasites have been enumerated in a table.

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